

A new tospovirus species infecting *Solanum esculentum* and *Capsicum annuum* in Thailand

P. Chiemsombat^A, M. Sharman^{B,C}, K. Srivilai^A, P. Campbell^B, D. Persley^B and S. Attathom^A

^ADepartment of Plant Pathology, Faculty of Agriculture at Kamphaeng Saen, Kasetsart University Kamphaeng Saen Campus, Nakhon Pathom 73140, Thailand.

^BDepartment of Employment, Economic Development and Innovation, 80 Meiers Road, Indooroopilly, Qld 4068, Australia.

^CCorresponding author. Email: Murray.Sharman@dpi.qld.gov.au

Abstract. Based on distinct sequences, serology and host range, isolates of a new tospovirus species infecting field crops of tomato and pepper in Thailand are shown to be members of the provisionally named Tomato necrotic ringspot virus.

Tospoviruses are among the most damaging and economically important group of plant viruses, causing significant crop losses in a wide range of ornamental and food crops in many regions of the world (Mumford *et al.* 1996). The greatest tospovirus diversity is found in the Asian region, with at least 14 species reported (Pappu *et al.* 2009). At least three species occur in Thailand including *Watermelon silver mottle virus* (WSMoV), Melon yellow spot virus (MYSV) and *Capsicum chlorosis virus* (CaCV) (Chiemsombat *et al.* 2008). Pepper (*Capsicum annuum*) and tomato (*Solanum esculentum*) displaying symptoms of bud necrosis and necrotic ringspots on leaves, stem and fruits were collected from production areas in Chiang Rai and Ratchaburi provinces of Thailand during 2007. This paper reports the molecular and biological identification of a new tospovirus species from these samples.

In March 2007 a symptomatic tomato sample (isolate-Q1510) was collected from the Ratchaburi region, while in December 2007 several symptomatic pepper samples were collected from the Chiang Rai region, one of which is hereafter referred to as isolate-CR. Field infected Q1510-tomato displayed symptoms of fine necrotic spots and rings on leaves (Fig. 1a), while pepper and tomato (Fig. 1b and c) inoculated with isolate-CR-infected pepper leaves, showed bud necrosis with severe mosaic and necrotic ring spots on leaves. A freeze-dried field leaf sample from isolate-Q1510 was negative by enzyme-linked immunosorbent assay (ELISA) for tospovirus serogroup IV (WSMoV/GBNV: AGDIA ELISA reagent set, Cat. No. SRA61500/0500) with $A_{405\text{ nm}}$ values comparable to those of the healthy controls while values of the freeze-dried positive control (CaCV) were 50 times greater than the means of the healthy controls.

Isolate-CR was transmitted by mechanical inoculation, using a field sample of pepper to tobacco (*Nicotiana tabacum* cv. Xanthi NC), *Physalis micrantha* and *Datura stramonium*, all of which produced severe mosaic symptoms. Isolate-CR was maintained in tomato cv. LY514 for subsequent tests. Diagnostic indicator

species for isolate-CR were shown to be *Vigna sinensis* cvv. Chiatai and KY Bush 693 with white local lesions at 7 days post-inoculation (Fig. 1d) while CaCV induced chlorotic concentric ring spots on the same host species (Chiemsombat *et al.* 2008). The test plants above were indexed by non-structural (NSs) gene-specific polymerase chain reaction (PCR), as described below, to confirm infection with isolate-CR.

Symptomatic leaf of isolate-CR-infected tomato reacted weakly in plate-trapped antigen (PTA) ELISA (Mowat and Dawson 1987) to polyclonal antibody A3 (PAb-A3) which was produced against expressed MYSV nucleoprotein (N) for detection of tospovirus serogroup IV species and MYSV (Gajanadana *et al.* 2006). This weak reaction against the PAb-A3 suggested the presence of tospovirus in these samples. Further field observations in 2009, with the use of PAb-A3 antibody in PTA-ELISA and inoculations to diagnostic species described below, indicated that several pepper crops in Nakhon Pathom province had significant losses due to bud necrosis disease caused by the same virus as isolate-CR (data unpublished).

For isolate-Q1510, RNA was extracted from leaf tissue using the Concert RNA Reagent (Invitrogen, Carlsbad, CA, US) before preparation of cDNA using SuperScript III reverse transcriptase (Invitrogen) and PCR using native *Taq* (Invitrogen) essentially as per the manufacturers' instructions. Degenerate primers gL3637 and gL4435c (Chu *et al.* 2001) were used to amplify an ~800 bp product of the partial L-gene which was directly sequenced. For amplification of the total medium RNA (mRNA) segment from isolate-Q1510, previously published tospovirus mRNA sequences (GenBank accessions: CaCV, NC_008303 and DQ256125; MYSV, NC_008307; WSMoV, NC_003841; *Groundnut bud necrosis virus* (GBNV), NC_003620; *Impatiens necrotic spot virus* (INSV), NC_003616 and DQ425095; and *Tomato spotted wilt virus* (TSWV), NC_002050, AY870390 and AY870389) were aligned and a degenerate primer TosMtotal (5' CCC GGG AGA GCA ATC

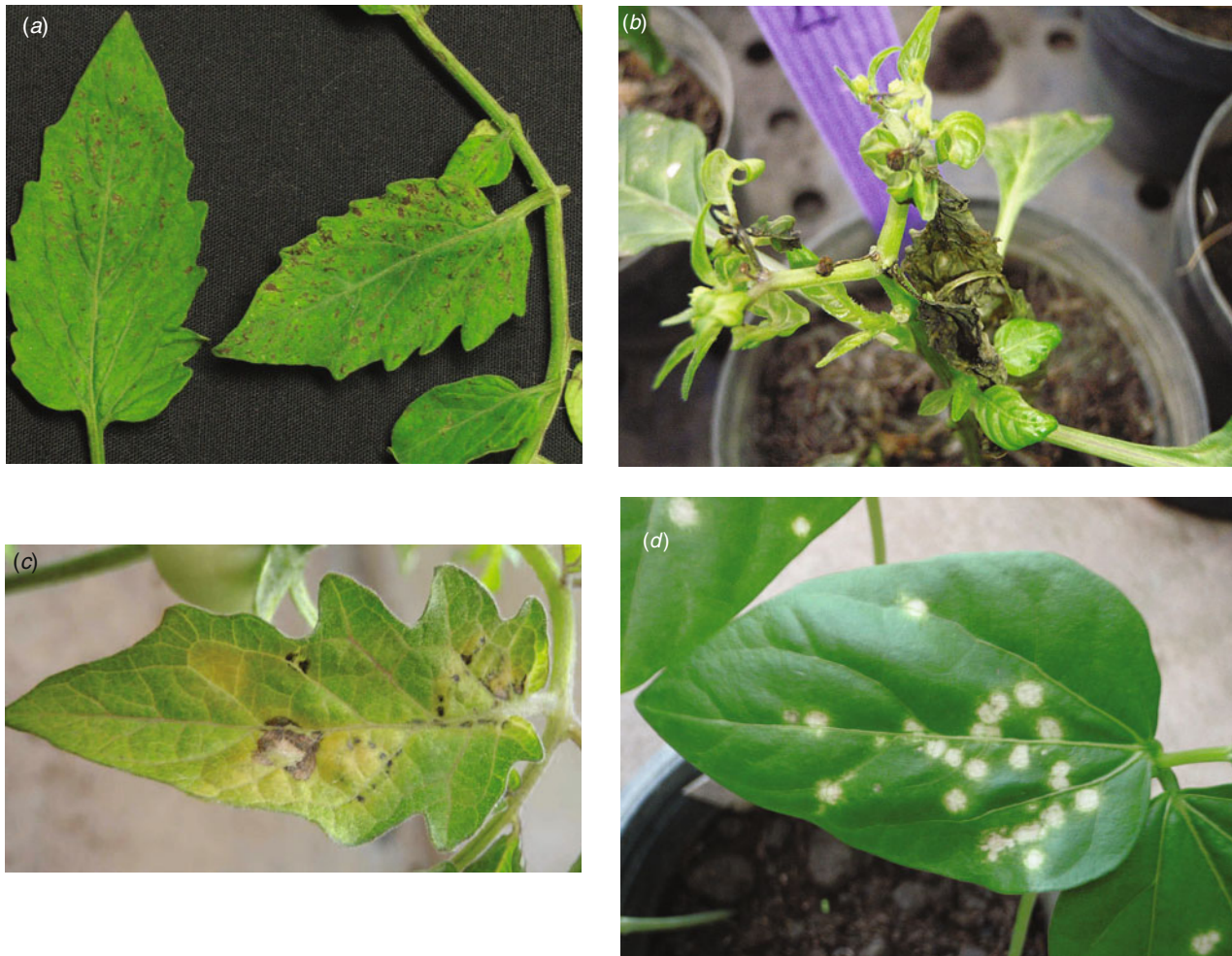


Fig. 1. (a) Symptoms of TNRV-Q1510 on field sample of tomato. Various hosts inoculated with TNRV-CR: (b) bud necrosis symptom on pepper; (c) necrotic ring spot leaves on tomato; and (d) *Vigna sinensis* cv. Chiatai displaying white local lesions.

RGT GC 3') designed to the complementary mRNA segment genome termini. The underlined sequence shown for primer TosMtotal above represents non-viral nucleotides added to increase primer stability in PCR. It was anticipated that the entire mRNA could be amplified using the TosMtotal primer. However, an internal region with homology to the primer sequence resulted in the amplification of an ~1300 bp product which was cloned into PCR 2.1 Topo vector (Invitrogen) and sequenced as above (GenBank HM136988).

For amplification of complete N-gene from the small RNA segment of isolate-Q1510, previously published sequences (GenBank accessions: WSMoV, NC_003843; GBNV, NC_003619; CaCV, DQ256123 and DQ355974; Iris yellow spot virus, AF001387; Calla lily chlorotic spot virus, AY867502; MYSV, NC_008300; Tomato yellow ring virus, DQ462163; and Tomato necrotic ringspot virus, FJ489600) were used to design primers. Primers TNRV.NPF (5' CAC TAT CTG ATA TAA ATG AT 3'), CS2821F (5' ACT TTG CAR ABC TGY TCA TA 3') and CS3477R (5' CGA CCA GAG CAA TCG AGG 3') were used to amplify two overlapping

products spanning 899 nt (GenBank HM113532) and containing the entire N-gene of 846 nt which encode a putative protein of 281 amino acids. PCR products were directly sequenced as described above.

For isolate-CR, RNA was extracted from leaf tissues using CTAB buffer followed by lithium chloride precipitation (Chang *et al.* 1993). Complete N-gene was amplified using the primers 3VGr4R (5' GTA AAC ACC ATG TCT AMC GT 3') and 5' WSMV Forward (5' GTG AGA GCA ACC CTG GAA CAG C 3') and complete NSs-gene was amplified using the primers 3VGr4R and 5VGr4F (5' TTA MAM TTC YAK MGA 3') (Chiemsoombat *et al.* 2008). Resulting PCR products were cloned into pGEMT-easy vector (Promega, Madison, WI, USA) and sequenced. For back-indexing of host range test plants, partial NSs gene-specific RT-PCR was used with the primers 3VGr4R and 5' WSMV Forward. The entire NSs gene was 1356 nt long. Isolate-CR sequences have been lodged with GenBank accession numbers FN806775 and FN677988 for N and NSs genes, respectively.

Table 1. Percentage identity of N protein amino acid sequences of selected tospoviruses taken from New and Old World clusters shown on phylogenetic tree in Fig. 2

Identity values for TNRV-Q1510 are shown and these are the same as those for TNRV-TT1 and TNRV-PP1

Virus	TNRV	WSMoV	CaCV	GBNV	CCSV	MYSV	IYSV	INSV	ZLCV	TSWV	PYSV
TNRV	–										
WSMoV	61	–									
CaCV	63	87	–								
GBNV	64	88	88	–							
CCSV	60	68	67	70	–						
MYSV	56	61	61	62	62	–					
IYSV	48	45	47	48	51	52	–				
INSV	27	31	32	30	26	29	30	–			
ZLCV	28	32	33	32	29	31	33	56	–		
TSWV	33	33	35	33	29	30	36	58	75	–	
PYSV	19	21	24	21	21	22	22	19	19	18	–

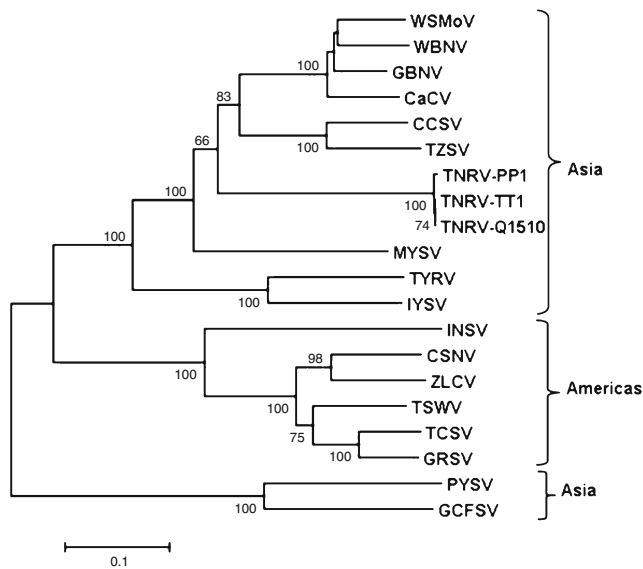


Fig. 2. Neighbour-joining dendrogram illustrating nucleoprotein (N) amino acid sequence relationships of tospovirus species. Horizontal distances are proportional to amino acid substitutions per site, and dendrograms were bootstrapped 500 times with branch values greater than 50% indicated. The two major geographical groupings of tospoviruses are as described by Pappu *et al.* (2009). Abbreviations and GenBank accession numbers used are: *Watermelon silver mottle virus* (WSMoV; NC_003843), *Capsicum chlorosis virus* (CaCV; FJ947157), *Groundnut bud necrosis virus* (GBNV; AY871098), *Watermelon bud necrosis virus* (WBNV; EU249351), *Calla lily chlorotic spot virus* (CCSV; AY867502), *Tomato zonate spot virus* (TZSV; NC_010489), *Tomato necrotic ringspot virus* (TNRV-Q1510, -TT1 and -PP1; HM113532, FJ489600 and FJ947153 respectively), *Melon yellow spot virus* (MYSV; NC_008300), *Tomato yellow ring virus* (TYRV; DQ462163), *Iris yellow spot virus* (IYSV; AF001387), *Impatiens necrotic spot virus* (INSV; NC_003624), *Chrysanthemum stem necrosis virus* (CSNV; AF067068), *Zucchini lethal chlorosis virus* (ZLCV; AF067069), *Tomato spotted wilt virus* (TSWV; NC_002051), *Tomato chlorotic spot virus* (TCSV; AF282982), *Groundnut ringspot virus* (GRSV; AF513219), *Groundnut yellow spot virus* (GYSV; AF013994) and *Groundnut chlorotic fan-spot virus* (GCFSV; AF080526).

The putative partial nucleocapsid (N) protein sequence of 248 amino acids for isolate-CR (GenBank FN806775) was 98% identical to isolate-Q1510 (GenBank HM113532) and to Tomato necrotic ringspot virus (TNRV) isolate-PP1, which was first recorded from Thailand pepper samples and provisionally named by other workers (GenBank FJ947153). The complete putative N protein sequence of 281 amino acids for isolate-Q1510 was 99.6% identical to TNRV-PP1 and 100% identical to TNRV-TT1 (GenBank FJ489600). As such, isolates-Q1510 and -CR are considered to be isolates of TNRV. The next most closely related tospovirus to TNRV-Q1510 was *Groundnut bud necrosis virus* (GBNV) with 64% identity for the complete N protein (Table 1). These results indicate TNRV is a distinct tospovirus species which clusters with the other Asian or Old World tospoviruses when comparing complete N protein sequences (Fig. 2). The putative complete

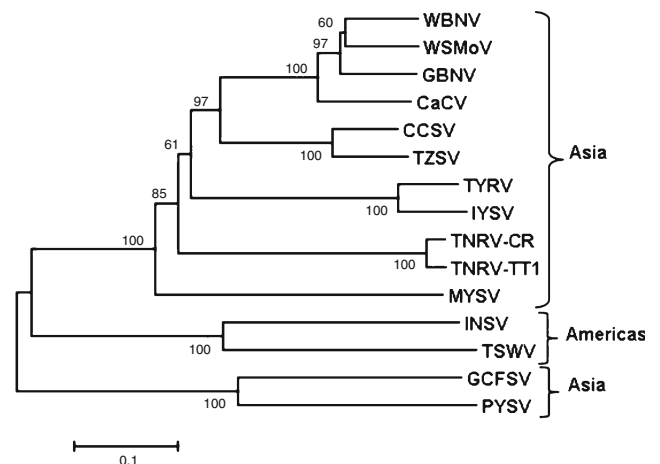


Fig. 3. Neighbour-joining dendrogram illustrating non-structural (NSs) protein amino acid sequence relationships of tospovirus species. Dendrogram construction, abbreviations and GenBank accession numbers as described for Fig. 2 plus TNRV-CR (GenBank FN677988).

NSs-gene amino acid sequences for isolates TNRV-CR and TNRV-TT1 (GenBank FN677988 and FJ489600 respectively) were 96% identical. Comparison of these TNRV isolates with other tospoviruses confirmed that TNRV is a new distinct tospovirus species (Fig. 3). The next most closely related tospovirus species to TNRV-CR was CCSV (GenBank AY867502) with only 55% identity for the complete NSs protein.

Partial mRNA nucleotide and amino acid sequence for TNRV-Q1510 were 99% and 98%, respectively, identical to TNRV-TT1 (GenBank FJ947152, Seepiban *et al.* 2009) over a 1230 nt (410 aa) overlap. The next closest related tospovirus to TNRV-Q1510 was GBNV (GenBank U42555) with 64% amino acid identity over a 410 aa overlap. Partial L RNA nucleotide sequence for TNRV-Q1510 (GenBank GQ487713, Sharman *et al.* 2009) was 99% identical, over a short overlap of 96 nt, to TNRV which has been reported from another tomato isolate from Thailand (GenBank GU479877) while the next closest related tospovirus was CCSV (GenBank FJ822962) with 78% identity over a 752 nt overlap.

This paper reports that isolates of a new tospovirus infecting field crops of tomato and pepper from different regions of Thailand are members of the provisionally named Tomato necrotic ringspot virus. Based on differences from other recognised tospovirus species with respect to serology, host range and nucleoprotein (N) and non-structural protein (NSs) sequence similarity, TNRV appears to be a distinct species grouping with other Old World species from Asia. The described differences to other tospovirus species satisfy the species demarcation criteria used by the International Committee on Taxonomy of Viruses (Fauquet *et al.* 2005). Many aspects of TNRV biology remain unclear and further studies on the alternative hosts, thrips vectors and the screening for natural resistance in germplasm collections will be important for development of effective management strategies. Recent advances in molecular techniques have now enabled the discovery of at least 19 distinct tospovirus species with one or more tospovirus species now found on every continent except Antarctica (Pappu *et al.* 2009). Along with TNRV, the number of tospovirus species described is likely to continue to increase with at least two further species being recently proposed (Ciuffo *et al.* 2009; Hassani-Mehraban *et al.* 2010).

References

- Chang S, Puryear J, Cairney J (1993) A simple and efficient method for isolating RNA from pine trees. *Plant Molecular Biology Reporter* **11**, 113–116. doi:10.1007/BF02670468
- Chiemsoombat P, Gajanandana O, Warin N, Hongprayoon R, Bhunchoth A, Pongsapich P (2008) Biological and molecular characterization of tospoviruses in Thailand. *Archives of Virology* **153**, 571–577. doi:10.1007/s00705-007-0024-3
- Chu F-H, Chao C-H, Chung M-H, Chen C-C, Yeh S-D (2001) Completion of the genome sequence of *Watermelon silver mottle virus* and utilization of degenerate primers for detecting tospoviruses in five serogroups. *Phytopathology* **91**, 361–368. doi:10.1094/PHYTO.2001.91.4.361
- Ciuffo M, Kurowski C, Vivoda E, Copes B, Masenga V, Falk BW, Turina M (2009) A New Tospovirus sp. in Cucurbit Crops in Mexico. *Plant Disease* **93**, 467–474. doi:10.1094/PDIS-93-5-0467
- Fauquet CM, Mayo MA, Maniloff J, Desselberger U, Ball LA (2005) 'Virus Taxonomy.' (Elsevier Academic Press: London)
- Gajanandana O, Seepiban C, Pansa-ard P, Bhunchoth A, Warin N, Himananto O, Chiemsoombat P, Pongsapich P, Hongprayoon R (2006) Production of antibodies for detection of tospoviruses in Thailand. In 'Proceedings of 44th Kasetsart University Annual Conference'. (Kasetsart University: Bangkok) pp. 445–451.
- Hassani-Mehraban A, Botermans M, Verhoeven J, Meekes E, Saaier J, Peters D, Goldbach R, Kormelink R (2010) A distinct tospovirus causing necrotic streak on *Alstroemeria* sp. in Colombia. *Archives of Virology* **155**, 423–428. doi:10.1007/s00705-010-0590-7
- Mowat WP, Dawson S (1987) Detection and identification of plant viruses by ELISA using crude sap extracts and unfractionated antisera. *Journal of Virological Methods* **15**, 233–247. doi:10.1016/0166-0934(87)90101-7
- Mumford RA, Barker I, Wood KR (1996) The biology of the tospoviruses. *The Annals of Applied Biology* **128**, 159–183. doi:10.1111/j.1744-7348.1996.tb07097.x
- Pappu HR, Jones RAC, Jain RK (2009) Global status of tospovirus epidemics in diverse cropping systems: successes achieved and challenges ahead. *Virus Research* **141**, 219–236. doi:10.1016/j.virusres.2009.01.009
- Seepiban C, Gajanandana O, Chiemsoombat P, Attathom T, Attathom S (2009) Characterization of Tomato necrotic ringspot virus (TNRV), a new tospovirus species isolated from Thailand. In 'Proceedings of Agricultural Biotechnology International Conference'. (Bangkok: Thailand)
- Sharman M, Chiemsoombat P, Campbell PR, Persley DM (2009) A new tospovirus species from Thailand. In 'Proceedings of the IX International Symposium on Thysanoptera and Tospoviruses'. (Gold Coast: Australia) 121pp.

Manuscript received 25 May 2010, accepted 1 July 2010