

## Emergence of serranid pigment abnormality syndrome (SPAS) in wire netting cod (*Epinephelus quoyanus*) from Heron Island on the southern Great Barrier Reef

B. K. Diggles<sup>A,D</sup>, I. Ernst<sup>B</sup> and S. Wesche<sup>C</sup>

<sup>A</sup>DigsFish Services, Banksia Beach, Qld 4507, Australia.

<sup>B</sup>Department of Agriculture and Water Resources, GPO Box 858, Canberra, ACT 2601, Australia.

<sup>C</sup>Queensland Department of Agriculture and Fisheries, GPO Box 46, Brisbane, Qld 4001, Australia.

<sup>D</sup>Corresponding author. Email: ben@digsfish.com

**Abstract.** Coral reefs worldwide are under increasing stress from anthropogenic impacts, but there are relatively few reports of increased rates of disease in coral reef fish. Herein we report the emergence of abnormal skin lesions in wild-caught wire netting cod (*Epinephelus quoyanus*) near Heron Island in the southern Great Barrier Reef. The lesion involves conspicuous darkening and disorganisation of the brown ‘wire netting’ colouration pattern typical of this species, most commonly on the lower jaw, premaxilla and head, with occasional involvement of the flanks and dorsal fin in some fish. The lesion was not present during research conducted in the mid-1990s; however, since it was first recorded in 2012, the prevalence of grossly visible lesions has increased to 16.9% in 2017, with fish >340 mm long most affected (prevalence 64.7%). These data suggest emergence of the lesion is a recent phenomenon and that causative factors may be age related. Abnormal pigmentation lesions have only been observed to affect *E. quoyanus* and coral trout (*Plectropomus leopardus*; since 2010). Given the species affected and the currently unknown aetiology of these lesions, we name the condition serranid pigment abnormality syndrome (SPAS). Further research is required to determine its geographic distribution, establish causation and describe the course of disease in *E. quoyanus*.

**Additional keywords:** chromatophoroma, coral reefs, disease, health, Serranidae, teleost.

Received 22 November 2017, accepted 6 February 2018, published online 17 May 2018

### Introduction

Baselines of ‘normal’ disease in wild fish populations remain elusive (Lafferty *et al.* 2004; Ward and Lafferty 2004), although it is often thought that emergence of new diseases may be an indicator of environmental stress (Harvell *et al.* 1999; Lafferty *et al.* 2004). It is well documented that coral reefs worldwide are under increased stress due to global warming (Couch *et al.* 2017; Hughes *et al.* 2017) and a range of anthropogenic stressors, including terrestrial run-off (McCulloch *et al.* 2003; Brodie *et al.* 2012; Wenger *et al.* 2016), overfishing (Cinner *et al.* 2009; Lamb *et al.* 2015) and tourism (Juhász *et al.* 2010; Lamb and Willis 2011; Au *et al.* 2014). On the Great Barrier Reef (GBR), these stressors have mostly been documented to affect corals in the form of bleaching events (Hughes *et al.* 2017), outbreaks of crown of thorns starfish (Fabricius *et al.* 2010; Wooldridge *et al.* 2015) and increased prevalence of coral diseases (Chen *et al.* 2017; Pollock *et al.* 2017).

In contrast, relatively few reports of disease outbreaks in wild fish or shellfish are found in the scientific literature on coral reefs. In the Caribbean, an acute large-scale epizootic in sea urchins (*Diadema antillarum*; see Lessios *et al.* 1984; Lessios 1988) preceded phase shifts towards algal dominance in coral

reefs in the early 1980s (Idjadi *et al.* 2010). Although a disease agent was never identified as the cause of the initial mortality event, a microbial pathogen was suspected (Bauer and Agerter 1987) and it has taken nearly 30 years before sea urchin populations (and corals) in the affected regions began to recover (Idjadi *et al.* 2010; Rodríguez-Barreras *et al.* 2015). Then, in 2000, spiny lobsters (*Panulirus argus*) in the Caribbean were found to be infected with a novel virus (*Panulirus argus* virus 1 (PaV1)) that caused disease and mortalities in juvenile lobsters (Behringer *et al.* 2011).

In teleosts, common naturally occurring parasites can occasionally cause disease in wild coral reef fish under certain conditions (Bunkley-Williams and Williams 1994; Landsberg 1995), whereas epizootics caused by bacterial disease agents, particularly *Streptococcus iniae* and *Streptococcus agalactiae*, have been reported in several countries (Ferguson *et al.* 2000; Bowater *et al.* 2012; Keirstead *et al.* 2014). Some populations of coral reef finfish near Hawaii have been described with skin diseases in the form of chromatophoromas (including melanomas) with unknown aetiology in butterflyfish (Okihira 1988) and surgeonfish (Work and Aeby 2014), whereas bicolor damselfish (*Stegastes partitus*) from Florida are affected by an

**Table 1. Serranid pigment abnormality syndrome (SPAS) prevalence and other data from field sampling of *Epinephelus quoyanus* from Heron Island between May 1996 and October 2017**Water temperature data are given as the mean  $\pm$  range for individual years and as the range for the period 2010–17. TL, total length

| Year    | Month     | Water temperature (°C) | Number of fish sampled | Mean TL (mm) | TL range (mm) | Number of fish $\geq$ 340 mm TL sampled | SPAS prevalence (%) | SPAS prevalence in fish $\geq$ 340 mm TL (%) |
|---------|-----------|------------------------|------------------------|--------------|---------------|---|---------------------|--|
| 1996    | May       | 23 $\pm$ 1             | 158                    | 295.4        | 230–370       | 19                                      | 0                   | 0  |
| 2010    | Nov.      | 25 $\pm$ 1             | 6                      | 321.7        | 295–360       | 2                                       | 0                   | 0  |
| 2011    | Oct.      | 24 $\pm$ 1             | 11                     | 301.3        | 270–360       | 2                                       | 0                   | 0  |
| 2012    | Oct.–Nov. | 24 $\pm$ 1             | 43                     | 306.4        | 172–370       | 11                                      | 2.33                | 9.1  |
| 2016    | Oct.      | 24 $\pm$ 1             | 64                     | 284.2        | 145–355       | 6                                       | 6.25                | 50   |
| 2017    | Oct.      | 25 $\pm$ 1             | 65                     | 306.8        | 210–380       | 17                                      | 16.92               | 64.7   |
| 2010–17 | Oct.–Nov. | 24–25                  | 189                    | 298.6        | 145–380       | 38                                      | 8.46                | 42.1   |

infectious viral disease (neurofibromatosis) involving development of neurofibromas and chromatophoromas (Schmale *et al.* 2002).

Herein we report for the first time the emergence of a pigmentation abnormality in the wire netting cod or longfin grouper (*Epinephelus quoyanus*) near Heron Island on the southern GBR. We began to encounter abnormally pigmented *E. quoyanus* from 2012 onwards as an incidental finding during unrelated research that required capture and tagging of *E. quoyanus* and stripey snapper (*Lutjanus carponotatus*) following on from research conducted in the same area in the mid-1990s (Diggles and Ernst 1997). Our awareness of the historical lack of pigment lesions in *E. quoyanus* and the recent emergence (since 2010) of superficially similar pigment lesions in another member of the Family Serranidae, namely the coral trout (*Plectropomus leopardus*), from the same region (Sweet *et al.* 2012; Lerebours *et al.* 2016) prompted us to publish these observations. Due to their presently unknown aetiology, we name these abnormalities serranid pigment abnormality syndrome (SPAS). This paper details the emergence of SPAS in the *E. quoyanus* population near Heron Island (Qld, Australia), describes the prevalence and gross pathology of the lesion and outlines the research that is needed to establish a better understanding of its geographical distribution, the course of disease and the mechanism(s) responsible for its emergence.

## Materials and methods

The present study was conducted in the Scientific Research Zone and Conservation Park zones of the Great Barrier Reef Marine Park in the waters adjacent to Heron Island (23°15'54"S, 151°32'53"E) in the southern section of the GBR. Examinations of *E. quoyanus* ( $n = 189$ ) were undertaken during five sampling trips between November 2010 and October 2017 (Table 1) and compared to historical data for *E. quoyanus* ( $n = 158$ ) obtained in May 1996 during a previous study (Diggles and Ernst 1997). Fish were captured from both reef flat (water depth <2 m) and reef edge (water depths 2–10 m) environments using rod-and-reel methods and artificial lures with barbless hooks, as described previously (Diggles and Ernst 1997). The anatomical hooking location of each fish was noted, its total length (TL) was measured to the nearest millimetre and this was followed by examination of each fish for gross signs of SPAS lesions or other

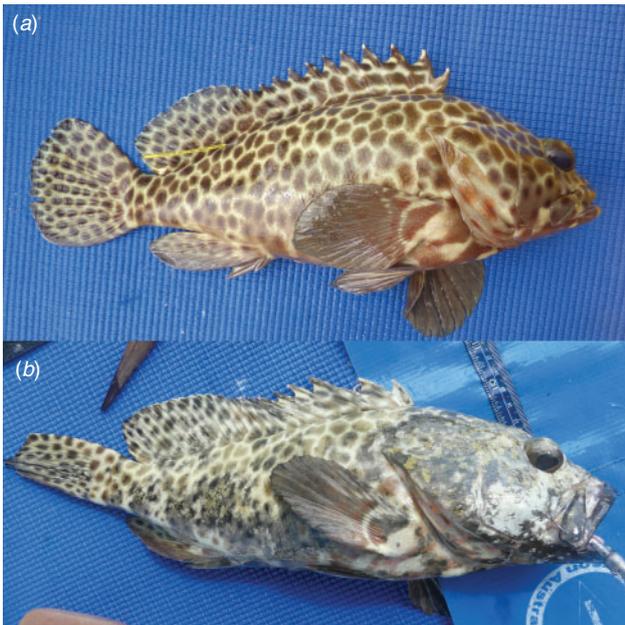
abnormalities. Each fish was then tagged with a plastic T-bar anchor tag with a unique identification number (TBA type; Hallprint, Hindmarsh Valley, SA, Australia) before being released at the site of capture. A smaller number of coral trout (*P. leopardus*;  $n = 60$ ) were incidentally captured using similar methods in the same locations in 2016 and 2017 and were subjected to measurement and same examination before being released at the site of capture without tags. These activities were conducted under Great Barrier Reef Marine Park Authority permit numbers G08/24996.1, G011/34020.1, G13/35806.1 and G15/37589.1.

To determine whether SPAS-affected fish were significantly larger than unaffected fish, Student's two-tailed *t*-tests were conducted on fish length using online statistical programs (<http://vassarstats.net/>, accessed 20 March 2018).

## Results

No pigment abnormalities were observed in any of the fish from the 1996, 2010 or 2011 studies, or in 42 of 43 fish examined in October and November 2012 (Fig. 1a). However, one fish (363 mm TL) captured on 1 November 2012 on the reef flat east of Heron Island was observed with conspicuous darkening and disorganisation of the brown 'wire netting' colouration pattern typical of this species over 100% of the head, 25–30% of the flanks and ~5% of the dorsal fin (Fig. 1b). To the naked eye, the lesions appeared smooth without obvious hyperplasia. Therefore, the sample prevalence for SPAS lesions in October and November 2012 was 1/43 = 2.33%, and, for all *E. quoyanus* captured since 1996 until that time, 1/218 = 0.46%.

In October 2016, an additional 64 *E. quoyanus* were examined, of which 4 (prevalence 6.25%) exhibited similar abnormally darkened pigmentation lesions without obvious hyperplasia most commonly involving the lower jaw, premaxilla and head (data not shown). Then, in October 2017, a further 65 *E. quoyanus* were examined, of which 11 (prevalence 16.92%) exhibited SPAS lesions most commonly on the lower jaw (prevalence 63.63%, mean coverage 52.14%), premaxilla (prevalence 36.36%, mean coverage 100%) and head (prevalence 72.72%, mean coverage 65.62%; Fig. 2), with involvement of the flanks (prevalence 45.45%, mean coverage 10–11%) and dorsal fin (prevalence 27.27%, mean coverage 6.67%; Fig. 3) in occasional fish (Table 2). After tagging and



**Fig. 1.** (a) An *Epinephelus quoyanus* exhibiting the normal colouration pattern for this species. (b) The first *E. quoyanus* (363-mm total length) observed with serranid pigment abnormality syndrome (SPAS) captured at a depth of <2 m on the reef flat east of Heron Island on 1 November 2012. Note the conspicuous darkening and disorganisation of the brown 'wire netting' colouration pattern over the head and flank.



**Fig. 2.** An *Epinephelus quoyanus* (350-mm total length) captured from the reef flat south-east of Heron Island on 8 October 2017. Note the serranid pigment abnormality syndrome (SPAS) lesion over nearly the entire head, including 100% of the lower jaw and maxillae.

release, none of the SPAS-affected fish were subsequently recaptured during these sampling trips, hence information on whether the lesions observed are progressive or resolve over time is currently not available.



**Fig. 3.** Left flank and dorsal fin of an *Epinephelus quoyanus* (351-mm total length) captured from the reef flat south-east of Heron Island on 8 October 2017. Note the serranid pigment abnormality syndrome (SPAS) lesion over the flank and rear dorsal fin involving darkening and shrinkage of the natural honeycomb pattern.

Only *E. quoyanus*  $\geq 325$  mm TL captured on the reef flat exhibited SPAS lesions, with prevalence of lesions increasing markedly with fish size above this threshold (Table 3). Mean sample prevalence for pigmentation lesions for all *E. quoyanus* captured between November 2010 and October 2017 ( $n = 189$ ) was  $16/189 = 8.47\%$  (Table 3), being highest in October 2017, with an overall prevalence of 16.92% and a prevalence of 64.7% in fish  $> 340$  mm TL during that month (Table 1). The mean size of SPAS-affected *E. quoyanus* ( $n = 16$ ; mean  $\pm$  s.e.m. size  $351 \pm 12$  mm TL, range 325–380 mm TL) was significantly greater than the mean size of unaffected fish ( $n = 173$ ; mean  $\pm$  s.e.m. size  $293.8 \pm 41.8$  mm TL, range 145–370 mm;  $P < 0.0001$ , Students two-tailed  $t$ -test). It was not possible to weigh fish at the point of capture, but one of the SPAS-affected fish examined in October 2017 was recorded in field notes as being noticeably underweight compared with clinically normal fish of the same size (Table 2).

The only other species of fish around Heron Island observed with SPAS-like pigmentation lesions were coral trout (*P. leopardus*). In a sample of 21 *P. leopardus* captured in 2016, 2 were SPAS affected (overall prevalence 9.5%; Table 4), with both SPAS-affected fish being larger fish ( $> 600$  mm fork length (FL)) sampled from deeper waters ( $> 10$  m) from the channel between Heron and Wistari reefs ( $n = 8$  fish sampled from the channel). In 2017, a sample of 39 *P. leopardus* captured from the channel, reef flat and reef edges near Heron Island revealed only 1 SPAS-affected fish (660 mm FL), which was caught and released from the reef edge (prevalence 2.56%; Fig. 4). The lesions on the coral trout were consistent in appearance with the melanised lesions reported by Sweet *et al.* (2012) to be present at a prevalence of 14.7% in the waters around Heron Island and nearby One Tree Island (Table 4).

Other than these two species, no abnormal pigmentation lesions were observed on any of the  $> 1000$  individual fish from  $> 17$  species captured and released by us during sampling trips

**Table 2.** Location and percentage coverage of serranid pigment abnormality syndrome (SPAS) lesions in individual *Epinephelus quoyanus* sampled from Heron Island in October 2017  
TL, total length

| Fish TL (mm)                         | Percentage coverage |            |       |            |             |            | Visually underweight |
|--------------------------------------|---------------------|------------|-------|------------|-------------|------------|----------------------|
|                                      | Lower jaw           | Premaxilla | Head  | Left flank | Right flank | Dorsal fin |                      |
| 360                                  | 100                 | 100        | 75    | 5          | 5           |            | No                   |
| 380                                  | 100                 | 100        | 100   | 15         | 15          |            | Yes                  |
| 350                                  |                     |            |       | 10         |             | 10         | No                   |
| 360                                  | 100                 | 100        | 70    | 10         | 10          | 5          | No                   |
| 351                                  | 5                   | 100        | 90    |            |             | 5          | No                   |
| 340                                  | 10                  |            |       |            |             |            | No                   |
| 355                                  |                     |            | 100   | 10         | 20          |            | No                   |
| 355                                  | 20                  |            |       |            |             |            | No                   |
| 350                                  |                     |            | 10    |            |             |            | No                   |
| 342                                  | 30                  |            | 70    |            |             |            | No                   |
| 353                                  |                     |            | 10    |            | 5           |            | No                   |
| Total number of affected individuals | 7/11                | 4/11       | 8/11  | 5/11       | 5/11        | 3/11       | 1/11                 |
| Mean coverage (%)                    | 52.14               | 100        | 65.62 | 10         | 11          | 6.67       | –                    |

**Table 3.** Prevalence of serranid pigment abnormality syndrome (SPAS) lesions in *Epinephelus quoyanus* examined at Heron Island between November 2010 and October 2017 versus fish total length

Only fish larger than 325 mm total length (TL) were affected, with the prevalence of SPAS increasing significantly with fish size

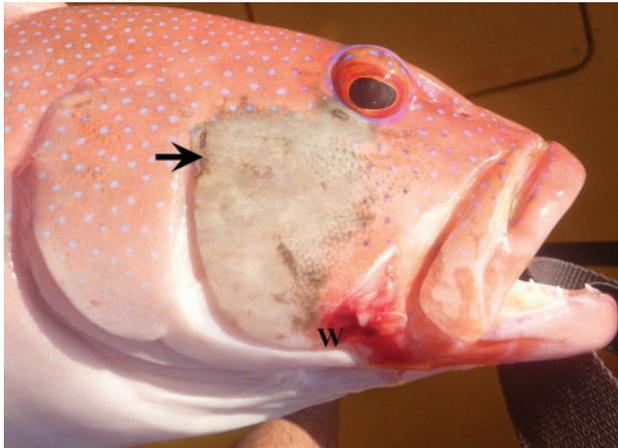
| Fish TL (mm)       | Number of normal fish | Number of fish with SPAS | Percentage of SPAS-affected fish in size class |
|--------------------|-----------------------|--------------------------|--|
| 0–150              | 2                     | 0                        | 0  |
| 151–200            | 2                     | 0                        | 0  |
| 201–250            | 17                    | 0                        | 0  |
| 251–300            | 72                    | 0                        | 0  |
| 301–320            | 34                    | 0                        | 0  |
| 321–350            | 35                    | 8                        | 18.60  |
| >350               | 11                    | 8                        | 42.11  |
| Number of all fish | 173                   | 16                       | 8.46   |
| Mean TL (mm)       | 293.8                 | 351.0 <sup>A</sup>       |  |

<sup>A</sup> $P < 0.0001$  (Student's two-tailed *t*-test).**Table 4.** Prevalence of serranid pigment abnormality syndrome (SPAS) lesions in coral trout (*Plectropomus leopardus*) sampled near Heron Island in October 2016 and 2017 compared with previously published dataWater temperature data are given as the mean  $\pm$  range for October 2016 and October 2017, and as the range for the period 2010–12. FL, fork length

| Year    | Month   | Water temperature (°C) | Number of fish sampled | FL range (mm) | SPAS prevalence (%) | Reference                  |
|---------|---------|------------------------|------------------------|---------------|---------------------|----------------------------|
| 2010–12 | –       | 21–27                  | 136                    | 344–639       | 14.7                | Sweet <i>et al.</i> (2012) |
| 2016    | October | 24 $\pm$ 1             | 21                     | 300–650       | 9.5                 | Present study              |
| 2017    | October | 25 $\pm$ 1             | 39                     | 295–660       | 2.56                | Present study              |

from 2010 onwards, including Lutjanidae (e.g. *Lutjanus carponotatus* ( $n > 300$ ) and *Lutjanus bohar* ( $n = 5$ )), Lethrinidae (e.g. *Lethrinus nebulosus* ( $n > 350$ ), *Lethrinus miniatus* ( $n > 150$ ), *Lethrinus atkinsoni* ( $n > 50$ ) and *Gymnocranius audleyi* ( $n = 10$ )), Labridae (e.g. *Cheilinus trilobatus* ( $n > 20$ ), *Choerodon cyanodus* ( $n > 20$ ) and *Choerodon venustus*

( $n > 30$ )), Carangidae (e.g. *Gnathanodon speciosus* ( $n > 5$ ) and *Caranx melampygus* ( $n = 1$ )) and other species of Serranidae (*Epinephelus fasciatus* ( $n > 50$ ), *Plectropomus maculatus* ( $n > 5$ ), *Epinephelus fuscoguttatus* ( $n = 2$ ), *Epinephelus cyanopodus* ( $n = 2$ ), *Cephalopholis miniata* ( $n = 5$ ) and *Variola louti* ( $n = 3$ )).



**Fig. 4.** An abnormal brown pigmented area (arrow) on the preopercle of a coral trout (*Plectropomus leopardus*) captured from the reef edge south-east of Heron Island. Similar pigmentation lesions were previously described from this species by Sweet *et al.* (2012). The haemorrhage in the mouth to the left of the maxilla is a hooking wound (W).

## Discussion

Evidence from baseline studies conducted in the mid-1990s (Diggles and Ernst 1997), combined with observations of gross pathology from the fish examined here, confirm that the SPAS lesions in *E. quoyanus* at Heron Island are new, abnormal and may have emerged some time after 2010. No lesions were observed between 1996 and 2011, with the first lesion observed by us in 2012.

Abnormal skin pigmentation lesions involving melanisation were reported previously by Sweet *et al.* (2012) in specimens of another serranid, namely *P. leopardus*, sampled from near Heron and One Tree islands in the southern GBR since 2010. In that study, 20 fish of the 136 sampled were affected (prevalence 14.7%), with no apparent relationship between fish size and intensity of the pigmentation lesions, which appeared to have some characteristics of tumours composed of melanophore pigment cells (melanophoroma; see Sweet *et al.* 2012). We also observed superficially similar pigmentation lesions in *P. leopardus* (Fig. 4), but at a lower prevalence (9.5% in 2016, 2.5% in 2017). Because *P. leopardus* was not a target species for our studies, our capture methods and locations differed from those of Sweet *et al.* (2012) and, because we were not able to conduct pathological investigations to confirm lesion identity, our sample prevalences for *P. leopardus* for 2016 and 2017 are not directly comparable to their data from 2010 to 2012. Therefore, no conclusions should be drawn regarding possible changes in the prevalence of pigment lesions in *P. leopardus* over time based on our data.

We can confirm the pigmentation lesions we first observed in *E. quoyanus* in 2012 have increased in prevalence over time and show a strong positive relationship with fish size, with no fish <325 mm TL being affected and a high prevalence of lesions occurring in fish >340 mm TL (up to 64.7% in October 2017).

Sweet *et al.* (2012) reported that *P. leopardus* with skin lesions ‘struck fishing hooks as strongly as healthy individuals, appeared to have good muscle tone and were assessed by

external examination as healthy aside from the skin discolouration’. The three affected *P. leopardus* examined by us in the present study also appeared, by gross observation and behaviour, to be in normal condition. Similarly, most SPAS-affected *E. quoyanus* captured in the present study appeared to have normal body condition, although one fish was in notably poor condition (based on visual assessment). It should be noted that capture methods used in the present study (line fishing with lures) could bias prevalence data (and the possible capture of any clinically affected fish) if SPAS-affected fish were more or less likely than unaffected fish to strike lures. However, because the same sampling methods were used for our original study (Diggles and Ernst 1997), their use here is entirely appropriate because it allows direct comparison of recent data with the original baseline data from 1996. Any future studies could examine the use of alternative or supplementary methods for examining lesion prevalence, such as underwater visual census.

The conditions of our research permit (for a different study) did not allow for destructive sampling of *E. quoyanus* for histopathological or other biological analysis (e.g. aging and sexing fish), hence the pathology of the lesion and risk factors remain to be determined. However, based on their darkened appearance, it is possible that increased melanin is a characteristic of the lesions in *E. quoyanus*, as was described for *P. leopardus* by Sweet *et al.* (2012) and Lerebours *et al.* (2016). Of the various pigment cell tumours in fish, melanophoromas (melanoma) are common relative to other types of tumours (Singaravel *et al.* 2017). Malignant neoplastic melanomas may be aggressively invasive and hyperplastic (Okiihiro 1988; Okiihiro *et al.* 1992; Raloff 2012; Work and Aeby 2014) compared with benign melanomas or preneoplastic melanosis, the latter being the most common skin pigmentation lesion recorded in redfish (*Sebastes mentella*) from the North Atlantic (Bogovski and Bakai 1989). The lesions in *E. quoyanus*, like those of *P. leopardus*, do not protrude from the skin surface and hence do not appear to be significantly invasive, hyperplastic or malignant; however, as discussed previously, further pathological characterisation is necessary. In any case, the emergence of pigment lesions in both *P. leopardus* and *E. quoyanus* (but apparently not other fish species) from the same region of the GBR at approximately the same time (2010–12), warrants further investigation to establish causation and examine the course of disease.

Because the lesions in *E. quoyanus* have only recently emerged, nothing is known in relation to risk factors that may contribute to their causation, except for fish length (and presumably age). Bogovski and Bakai (1989) found that melanosis and melanophoromas in *S. mentella* were more common in certain regions of the North Atlantic, were positively correlated with fish size (only occurring in fish >26 cm, with prevalence exceeding 50% in fish >40 cm) and exhibited variation between sexes. In that study, pigmented lesions occurred in females two- to fivefold more often than in males, whereas in male fish 84% of pigment lesions occurred on the head and caudal fin (Bogovski and Bakai 1989). In *E. quoyanus* at Heron Island in October 2017, SPAS lesions were only observed on fish >340 mm TL, most commonly on the lower jaw (prevalence 63.63%) and head (prevalence 72.72%; Table 2). Exposure to

ultraviolet (UV) light was one of the potential risk factors discussed by Sweet *et al.* (2012) for pigmentation lesions in *P. leopardus*, and this could also be a risk factor for *E. quoyanus* given their shallow water (<2 m) habitat. In contrast, the melanised pigment lesions in *S. mentella* were unlikely to be related to UV exposure because those fish were taken in bottom or mid-water trawls at depths of 60–420 m, although the lesions were more common in fish taken at ‘shallower’ depths of 60–250 m (Bogovski and Bakai 1989).

Poor water quality is known to be associated with non-infectious diseases of fish, including neoplasms (Kinae *et al.* 1990). Recent water quality studies in Queensland have shown that biologically relevant reductions in water quality occur up to 100 km offshore during episodic flood events (Brodie *et al.* 2012). Heron Island is ~70 km offshore from mainland Queensland, and hence is within the range of exposure of flood plumes. However, Bogovski and Bakai (1989) sampled fish from the mid-Atlantic and suggested that melanised pigment lesions in *S. mentella* were not likely to be related to declines in water quality but due to aging-related factors (senescence) or reduced gene flow within certain fish populations. Reduced gene flow was also suggested by Sweet *et al.* (2012), although infectious aetiologies are also possible (Schmale *et al.* 2002; Work and Aebly 2014).

Fish length (and possibly age) is an important risk factor for SPAS in *E. quoyanus*, because lesion prevalence increases markedly once total fish length exceeds 325 mm. The maximum size of *E. quoyanus* is between 380 and 400 mm (Fishbase 2017), but the minimum legal size of *E. quoyanus* in Queensland is 380 mm. Although the fish population sampled at Heron Island during the present study exists in a research zone where the taking of fish without scientific permits is prohibited, this species is effectively protected throughout Queensland by fisheries regulations, and it is possible that similar size and age structures of *E. quoyanus* populations will occur outside marine park no-take areas closed to fishing. The occurrence of SPAS lesions in *E. quoyanus* populations beyond Heron Reef would require surveys designed to elucidate the geographic extent of this syndrome.

Given that SPAS lesions are more prevalent in larger fish, it is possible that these lesions are gross signs of senescence or a progressive disease. Further research is required to describe the course of disease in *E. quoyanus*, to establish causation and ascertain whether the condition is infectious. This could be done in wild fish by sampling affected fish from various locations to obtain pathological samples, genetic information and age at length and sex data. Tagging and recapturing of affected fish would allow temporal monitoring of the extent of the lesions to determine whether they resolve or progress and increase in severity or the area of affected skin over time. However, if natural mortality rates or tag loss rates are high, such an approach may not yield enough information, which would necessitate bringing affected fish into captivity to monitor whether the disease progresses over time under controlled conditions.

### Conflicts of interest

The authors declare that they have no conflicts of interest.

### Acknowledgements

The authors thank N. Diggles and D. Evans for assistance with the collection of specimens in the field, the staff and management of Heron Island Research Station for the use of their facilities and two anonymous referees who provided valuable suggestions on earlier drafts of the manuscript.

### References

- Au, A. C., Zhang, L., Chung, S., and Qiu, J.-W. (2014). Diving associated coral breakage in Hong Kong: differential susceptibility to damage. *Marine Pollution Bulletin* **85**, 789–796. doi:10.1016/J.MARPOLBUL.2014.01.024
- Bauer, J. C., and Agerter, C. J. (1987). Isolation of bacteria pathogenic for the sea urchin *Diadema antillarum* (Echinodermata: Echinoidea). *Bulletin of Marine Science* **40**, 161–165.
- Behringer, D. C., Butler, M. J., Shields, J. D., and Moss, J. (2011). Review of *Panulirus argus* virus1 – a decade after its discovery. *Diseases of Aquatic Organisms* **94**, 153–160. doi:10.3354/DAO02326
- Bogovski, S. P., and Bakai, Y. I. (1989). Chromatoblastomas and related pigmented lesions in deepwater redbfish, *Sebastes mentella* (Travin), from North Atlantic areas, especially the Irminger Sea. *Journal of Fish Diseases* **12**, 1–13. doi:10.1111/J.1365-2761.1989.TB01234.X
- Bowater, R. O., Forbes-Faulkner, J., Anderson, I. G., Condon, K., Robinson, B., Kong, F., Gilbert, G. L., Reynolds, A., Hyland, S., McPherson, G., Brien, J. O., and Blyde, D. (2012). Natural outbreak of *Streptococcus agalactiae* (GBS) infection in wild giant Queensland grouper, *Epinephelus lanceolatus* (Bloch), and other wild fish in northern Queensland, Australia. *Journal of Fish Diseases* **35**, 173–186. doi:10.1111/J.1365-2761.2011.01332.X
- Brodie, J. E., Kroon, F. J., Schaffelke, B., Wolanski, E. C., Lewis, S. E., Devlin, M. J., Bohnet, I. C., Bainbridge, Z. T., Waterhouse, J., and Davis, A. M. (2012). Terrestrial pollutant runoff to the Great Barrier Reef: An update of issues, priorities and management responses. *Marine Pollution Bulletin* **65**, 81–100. doi:10.1016/J.MARPOLBUL.2011.12.012
- Bunkley-Williams, L., and Williams, E. H. Jr (1994). Diseases caused by *Trichodina spheroidesi* and *Cryptocaryon irritans* (Ciliophora) in wild coral reef fishes. *Journal of Aquatic Animal Health* **6**, 360–361. doi:10.1577/1548-8667(1994)006<0360:DCBTSA>2.3.CO;2
- Chen, C. C. M., Bourne, D. G., Drovandi, C. C., Mengersen, K., Willis, B. L., Caley, M. J., and Sato, Y. (2017). Modelling environmental drivers of black band disease outbreaks in populations of foliose corals in the genus *Montipora*. *PeerJ* **5**, e3438. doi:10.7717/PEERJ.3438
- Cinner, J. E., McClanahan, T. R., Graham, N. A., Pratchett, M. S., Wilson, S. K., and Raina, J. B. (2009). Gear-based fisheries management as a potential adaptive response to climate change and coral mortality. *Journal of Applied Ecology* **46**, 724–732. doi:10.1111/J.1365-2664.2009.01648.X
- Couch, C. S., Burns, J. H. R., Liu, G., Steward, K., Gutlay, T. N., Kenyon, J., Eakin, C. M., and Kosaki, R. K. (2017). Mass coral bleaching due to unprecedented marine heatwave in Papahānaumokuākea Marine National Monument (Northwestern Hawaiian Islands). *PLoS One* **12**, e0185121. doi:10.1371/JOURNAL.PONE.0185121
- Diggles, B. K., and Ernst, I. (1997). Hooking mortality of two species of shallow-water reef fish caught by recreational angling methods. *Marine and Freshwater Research* **48**, 479–483. doi:10.1071/MF96108
- Fabricius, K. E., Okaji, K., and De'ath, G. (2010). Three lines of evidence to link outbreaks of the crown-of-thorns seastar *Acanthaster planci* to the release of larval food limitation. *Coral Reefs* **29**, 593–605. doi:10.1007/S00338-010-0628-Z
- Ferguson, H. W., St. John, V. S., Roach, C. J., Willoughby, S., Parker, C., and Ryan, R. (2000). Caribbean reef fish mortality associated with *Streptococcus iniae*. *The Veterinary Record* **147**, 662–664.
- Fishbase (2017). *Epinephelus quoyanus* (Valenciennes, 1830), longfin grouper. Available at <http://www.fishbase.org/summary/6475> [Verified 20 March 2018].

- Harvell, C. D., Kim, K., Burkholder, J. M., Colwell, R. R., Epstein, P. R., Grimes, D. J., Hofmann, E. E., Lipp, E. K., Osterhaus, A. D., Overstreet, R. M., Porter, R. W., Smith, G. W., and Vasta, G. R. (1999). Emerging marine diseases – climate links and anthropogenic factors. *Science* **285**, 1505–1510. doi:10.1126/SCIENCE.285.5433.1505
- Hughes, T. P., Kerry, J. T., Álvarez-Noriega, M., Álvarez-Romero, J. G., Anderson, K. D., Baird, A. H., Babcock, R. C., Beger, M., Bellwood, B. R., Berkelmans, R., Bridge, T. C., Butler, I. R., Byrne, M., Cantin, N. E., Comeau, S. E., Connolly, S. R., Cumming, G. S., Dalton, S. J., Diaz-Pulido, G., Eakin, C. M., Figueira, W. F., Gilmour, J. P., Harrison, H. B., Heron, S. F., Hoey, A. S., Hobbs, J. A., Hoogenboom, M. O., Kennedy, E. V., Kuo, C., Lough, J. M., Lowe, R. J., Liu, G., McCulloch, M. T., Malcolm, H. A., McWilliam, M. J., Pandolfi, J. M., Pears, R. J., Pratchett, M. S., Schoepf, V., Simpson, T., Skirving, W. J., Sommer, B., Torda, G., Wachenfeld, D. R., Willis, B. L., and Wilson, S. K. (2017). Global warming and recurrent mass bleaching of corals. *Nature* **543**, 373–377. doi:10.1038/NATURE21707
- Idjadi, J. A., Haring, R. N., and Precht, W. F. (2010). Recovery of the sea urchin *Diadema antillarum* promotes scleractinian coral growth and survivorship on shallow Jamaican reefs. *Marine Ecology Progress Series* **403**, 91–100. doi:10.3354/MEPS08463
- Juhasz, A., Ho, E., Bender, E., and Fong, P. (2010). Does use of tropical beaches by tourists and island residents result in damage to fringing coral reefs? A case study in Moorea French Polynesia. *Marine Pollution Bulletin* **60**, 2251–2256. doi:10.1016/J.MARPOLBUL.2010.08.011
- Keirstead, N. D., Brake, J. W., Griffin, M. J., Halliday-Simmonds, I., Thrall, M. A., and Soto, E. (2014). Fatal septicemia caused by the zoonotic bacterium *Streptococcus iniae* during an outbreak in Caribbean reef fish. *Veterinary Pathology* **51**, 1035–1041. doi:10.1177/0300985813505876
- Kinae, N., Yamashita, M., Tomita, I., Kimura, I., Ishida, H., Kumai, H., and Nakamura, G. (1990). A possible correlation between environmental chemicals and pigment cell neoplasia in fish. *The Science of the Total Environment* **94**, 143–153. doi:10.1016/0048-9697(90)90369-6
- Lafferty, K. D., Porter, J. W., and Ford, S. E. (2004). Are diseases increasing in the ocean? *Annual Review of Ecology Evolution and Systematics* **35**, 31–54. doi:10.1146/ANNUREV.ECOLSYS.35.021103.105704
- Lamb, J. B., and Willis, B. L. (2011). Using coral disease prevalence to assess the effects of concentrating tourism activities on offshore reefs in a tropical marine park. *Conservation Biology* **25**, 1044–1052. doi:10.1111/J.1523-1739.2011.01724.X
- Lamb, J. B., Williamson, D. H., Russ, G. R., and Willis, B. L. (2015). Protected areas mitigate diseases of reef building corals by reducing damage from fishing. *Ecology* **96**, 2555–2567. doi:10.1890/14-1952.1
- Landsberg, J. H. (1995). Tropical reef fish disease outbreaks and mass mortalities in Florida, USA: what is the role of dietary biological toxins. *Diseases of Aquatic Organisms* **22**, 83–100. doi:10.3354/DAO022083
- Lerebours, A., Chapman, E. C., Sweet, M. J., Heupel, M. R., and Rotchell, J. M. (2016). Molecular changes in skin pigmented lesions of the coral trout *Plectropomus leopardus*. *Marine Environmental Research* **120**, 130–135. doi:10.1016/J.MARENRES.2016.07.009
- Lessios, H. A. (1988). Mass mortality of *Diadema antillarum* in the Caribbean: what have we learned? *Annual Review of Ecology and Systematics* **19**, 371–393. doi:10.1146/ANNUREV.ES.19.110188.002103
- Lessios, H. A., Cubit, J. D., Robertson, D. R., Shulman, M. J., Parker, M. R., Garrity, S. D., and Levings, S. C. (1984). Mass mortality of *Diadema antillarum* on the Caribbean coast of Panama. *Coral Reefs* **3**, 173–182. doi:10.1007/BF00288252
- McCulloch, M., Fallon, S., Wyndham, T., Hendy, E., Lough, J., and Barnes, D. (2003). Coral record of increased sediment flux to the inner Great Barrier Reef since European settlement. *Nature* **421**, 727–730. doi:10.1038/NATURE01361
- Okiihiro, M. S. (1988). Chromatophoromas in two species of Hawaiian butterflyfish, *Chaetodon multicinctus* and *C. miliaris*. *Veterinary Pathology* **25**, 422–431. doi:10.1177/030098588802500604
- Okiihiro, M. S., Whipple, J. A., Groff, J. M., and Hinton, D. E. (1992). Chromatophoromas and related hyperplastic lesions in Pacific rockfish (*Sebastes* spp.). *Marine Environmental Research* **34**, 53–57. doi:10.1016/0141-1136(92)90082-W
- Pollock, F. J., Wada, N., Torda, G., Willis, B. L., and Bourne, D. G. (2017). Repeated sampling of white syndrome-affected corals reveals distinct microbiome at disease lesion fronts. *Applied and Environmental Microbiology* **83**(2), e02799-16. doi:10.1128/AEM.02799-16
- Raloff, J. (2012). Epidemic of skin lesions reported in reef fish. In *Science News*, 1 August 2012. (Society for Science & the Public.) Available at <https://www.sciencenews.org/blog/science-public/epidemic-skin-lesions-reported-reef-fish> [Verified 20 March 2018].
- Rodriguez-Barreras, R., Durán, A., López-Morella, J., and Sabat, A. M. (2015). Effect of fish removal on the abundance and size structure of the sea urchin *Diadema antillarum*: a field experiment. *Marine Biology Research* **11**, 1100–1107. doi:10.1080/17451000.2015.1064140
- Schmale, M. C., Gibbs, P. D. L., and Campbell, C. E. (2002). A virus-like agent associated with neurofibromatosis in damselfish. *Diseases of Aquatic Organisms* **49**, 107–115. doi:10.3354/DAO049107
- Singaravel, V., Gopalakrishnan, A., and Raja, K. (2017). Iridophoroma in Indian mackerel, *Rastrelliger kanagurta* (Cuvier, 1816). *Journal of Applied Ichthyology* **33**, 116–118. doi:10.1111/JAI.13226
- Sweet, M., Kirkham, N., Bendall, M., Currey, L., Bythell, J., and Heupel, M. (2012). Evidence of melanoma in wild marine fish populations. *PLoS One* **7**, e41989. doi:10.1371/JOURNAL.PONE.0041989
- Ward, J. R., and Lafferty, K. D. (2004). The elusive baseline of marine disease: are diseases in ocean ecosystems increasing? *PLoS Biology* **2**, e120. doi:10.1371/JOURNAL.PBIO.0020120
- Wenger, A. S., Williamson, D. H., da Silva, E. T., Ceccarelli, D. M., Browne, N. K., Petus, C., and Devlin, M. J. (2016). Effects of reduced water quality on coral reefs in and out of no-take marine reserves. *Conservation Biology* **30**, 142–153. doi:10.1111/COBI.12576
- Wooldridge, S. A., Brodie, J. E., Kroon, F. J., and Turner, R. D. (2015). Ecologically based targets for bioavailable (reactive) nitrogen discharge from the drainage basins of the Wet Tropics region, Great Barrier Reef. *Marine Pollution Bulletin* **97**, 262–272. doi:10.1016/J.MARPOLBUL.2015.06.007
- Work, T. M., and Aeby, G. S. (2014). Skin pathology in Hawaiian goldring surgeonfish, *Ctenochaetus strigosus* (Bennett). *Journal of Fish Diseases* **37**, 357–362. doi:10.1111/JFD.12112