

In-vitro activity of ceftiofur against Australian isolates of the family *Pasteurellaceae* associated with respiratory disease in cattle and pigs

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Ceftiofur is a broad spectrum, b-lactamase-resistant cephalosporin (Yancey *et al* 1987). Studies performed outside Australia have established that ceftiofur inhibits the growth of isolates of *Pasteurella multocida*, *Pasteurella haemolytica*, *Actinobacillus pleuropneumoniae*, *Haemophilus somnus*, *Escherichia coli* and *Salmonella typhimurium* at concentrations of 0.25 mg/ml or less (Yancey *et al* 1987; Post *et al* 1991; Watts *et al* 1994).

A field study in Canada has demonstrated that ceftiofur is effective in the treatment of natural outbreaks of respiratory disease in feedlot cattle proving superior to trimethoprim-sulfadoxine (Jim *et al* 1992). In that study, cattle receiving ceftiofur had a significantly lower percentage of animals requiring more than three days of treatment for the initial treatment episode compared with the cattle receiving trimethoprim-sulfadoxine.

In this paper, we report on the *in vitro* antibacterial activity of ceftiofur against isolates of *P. multocida* and *P. haemolytica* (19 and 34 respectively) obtained from Australian cattle and isolates of *P. multocida* and *A. pleuropneumoniae* (30 and 50 respectively) from Australian pigs. The *Pasteurella* isolates were selected from as diverse a background as possible with isolates from New South Wales (1 *P. haemolytica*, 11 bovine *P. multocida* and 10 porcine *P. multocida* isolates), Queensland (33 *P. haemolytica*, 7 bovine *P. multocida* and 11 porcine *P. multocida* isolates) and Victoria (1 bovine *P. multocida* isolate and 9 porcine *P. multocida* isolates). The isolates of *A. pleuropneumoniae* examined were selected to represent the known genetic diversity of this species in Australia (Hampson *et al* 1993) and included all 7 serovars known to occur in Australia (Hampson *et al* 1993; Blackall and Pahoff 1995). Representatives of the currently unserotypable isolates were also included (Blackall and Pahoff 1995). A reference strain of *E. coli* (ATCC25922) was used as a control strain.

The minimal inhibitory concentration (MIC) of ceftiofur* for the isolates was determined by a standardised agar dilution method as described previously (Stephens *et al* 1993) with some modifications. Firstly, the antibiotic plates did not contain lysed horse blood. Secondly, for the *A. pleuropneumoniae* isolates, all media were supplemented with 0.0024% (w/v) reduced nicoti-

namide adenine dinucleotide (NADH). The agar plates contained a final concentration of ceftiofur that varied from 50 mg/ml to 0.20 mg/ml. The test organisms were grown overnight in tryptose phosphate broth† for the *Pasteurella* spp or tryptose phosphate broth† plus 0.0025% NADH for *A. pleuropneumoniae* and were diluted 10⁻³, to yield about 5 x 10⁷ colony forming units/mL (Blackall *et al* 1995). One 1 µL of this dilution was then inoculated onto the plates. A plate that did not contain any antibiotic was included with each set of organisms. All organisms were also plated onto 5% sheep blood agar to confirm the purity of the inoculum. Each plate was also inoculated with the control strain of *E. coli* (ATCC25922). The inoculated plates were incubated aerobically at 37°C for 18 hours. The MIC was taken as the lowest concentration to prevent growth. The MIC₉₅ was defined as that concentration of ceftiofur that inhibited 95% of the isolates examined.

The MIC for the reference strain of *E. coli* was always 0.78 mg/mL. This value is within the expected range (0.25 - 1.0 mg/mL) for this strain (Anon 1990). The MICs for all the bovine and porcine *P. multocida*, the bovine *P. haemolytica* and the *A. pleuropneumoniae* isolates were ≤ 0.2 mg/mL. As no isolate of any of the three species tested was capable of growing at the minimum antibiotic concentration used in this study (0.2 mg/mL), the MIC₉₅ of ceftiofur for all the isolates examined in this study was ≤ 0.2 mg/mL. The recommended *in vitro* interpretative criteria for ceftiofur are as follows: growth at ≥ 8 mg/mL = resistant, growth at 4 mg/mL but no higher = intermediate sensitivity; growth at 2 mg/mL but no higher = susceptible. On the basis of these figures, all 34 isolates of *P. haemolytica*, all 49 isolates of *P. multocida* and all 50 isolates of *A. pleuropneumoniae* were susceptible to ceftiofur.

Similar studies, based on isolates of these species obtained from American animals, reported an MIC₉₅ of ceftiofur for these three species of either ≤ 0.06 mg/ml (Yancey *et al* 1987; Watts *et al* 1994) or ≤ 0.125 mg/mL (Post *et al* 1991). The higher figure reported in our study is explained by the fact that the lowest antibiotic dilution used in our study was 0.2 mg/ml. In both our study and the three previous studies (Yancey *et al* 1987; Post *et al* 1991; Watts *et al* 1994), all isolates examined have been found to be susceptible to ceftiofur. The results of our study have provided evidence that ceftiofur is highly effective *in vitro* against Australian bovine and porcine isolates of *Pasteurella* spp and porcine isolates of *A. pleuropneumoniae*.

The assistance of Dr M White (Allied Feeds), Ms M Adamson (Victorian Department of Agriculture) and Mr P Duffy (Queensland Department of Primary Industries) in providing some of the isolates used in this study is gratefully acknowledged.

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(Accepted for publication 27 October 1995)

* Ceftiofur sodium, Lot No. 555-1212, Upjohn Company, Kalamazoo, USA

† Oxoid CM283, Oxoid Australia Pty Ltd, West Heidelberg, Vic