

Impacts of pesticides and fertilisers on soil biota.

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Introduction

The effects of inorganic amendments (fertilisers and pesticides) on soil biota that are reported in the scientific literature are, to say the least, variable. Though there is clear evidence that certain products can have significant impacts, the effects can be positive or negative. This is not surprising when you consider the number of organisms and amount of different functional groups, the number of products and various rates at which they could be applied, the methods of application and the environmental differences that occur in soil at a micro scale (within centimetres) in a paddock, let alone between paddocks, farms, catchments, regions etc. It therefore becomes extremely difficult to draw definitive conclusions from the reported results in order to summarise the impacts of these inputs. Several research trials and review papers have been published on this subject and most similarly conclude that the implications of many of the effects are still uncertain.

Types of effects

Effects of fertilisers and pesticides can be either direct (immediate or short-term impacts) due to harm to the organisms that come in contact with the chemical, or indirect due to changes caused by the chemical to the environment and/or food source of the organisms being studied. Time of sampling (in relation to last rainfall event, soil temperature, prevalence of food source, background nutrient levels in soil, soil pH), can be pivotal in the outcome of results as soil environmental conditions impinge greatly on microbial activity. Other factors include time between fertiliser/pesticide application and sampling, formulation and rate of fertilisers, depth of soil sampled, effects on food sources i.e. impacts on predator-prey interactions.

In some cases there maybe a short-term but no long-term effect due to resilience of soil biological populations (ability to come back with time) (Angus *et al.* 1999). Altering the detritus food web through the use of broad-spectrum pesticides reduces biological diversity and therefore alters the balance or equilibrium of the ecosystem. This could lead to pest resurgence through the absence of natural predators (Wildermuth *et al.* 1997).

Fertilisers

The direct effects of fertilisers on soil biota can be short- (obvious in the first season after application of the fertiliser) or long-term (if repeated additions are required to see the effect). Indirect effects are usually long-term (take more than one season to develop) and are due to changes in pH or changes in productivity, residue inputs and soil organic matter levels (Bunemann and McNeill 2004). These effects become important in agriculture when nutrient availability to plants and hence crop productivity are changed due to the effect.

One issue generally of concern to growers has been the potential damage to soil microorganisms from high concentrations of ammonia fertiliser applied in bands. Toxic effects of ammonia on soil organisms are known but injection of anhydrous ammonia into soils has impacts only in the zone/band of application and the effects are usually short-term. A project conducted in the mid 1990s in southern NSW showed that the injection of ammonia and urea in bands reduced total microbial activity in the narrow band of application for a period of 5 weeks, after which levels returned to normal. Recovery was however a little slower for some groups of organisms. There was an 80% reduction in the number of protozoa and their numbers did not return to normal after 5 weeks. Also, there was a large increase in the number of nitrifying bacteria in the soil five weeks after application (Angus *et al.* 1999). The researchers concluded that most groups of soil microorganisms were not adversely affected by banded ammonia or urea with a single year of applications however they could not draw any conclusions about long-term effects on soil microorganisms.

Work by Biederbeck *et al.* (1996) in Canada showed minimal impact on soil microbial populations and soil quality after ten years of fertilisation with urea and anhydrous ammonia. However, Stirling (pers. comm.) found that long-term use (9 years) of urea or ammonium fertilisers in a trial near Tullooona, NSW, had impacts on the longer lifecycle omnivorous nematodes. Numbers were lower in plots fertilised at 90 kg N/ha than in those not fertilised in soil from 5-15 and 15-30 cm depth intervals. These effects were still evident after the growing of a wheat crop. Other shorter life-cycle nematodes were not affected. Measurements of microbial biomass and activity on the same soil taken down to 30 cm in soil in the same trial were similarly not significantly different due to different fertiliser treatments (Van Zwieten pers. comm.).

In a review on the effects of management practices on soil mesofauna (protozoa, amoeba and nematodes), Gupta (1994) compares reported effects of N fertiliser on protozoa. The diverse range of responses illustrates dangers in extrapolating from one data set in one environment. Effects reported covered significant increases; initial decreases that then became increases in a short period of time; and stabilisation of numbers when lime was added with fertiliser. Similarly, effects of fertiliser applications on nematode populations were diverse and changes in both abundance and trophic diversity have been reported (see Gupta 1994).

Mycorrhizal fungi have been consistently reported as being decreased by P fertiliser but the extent to which this occurs may be dependent on species of fungus involved and level of plant available P in the soil. Phosphorus applications of 50 mg.kg⁻¹ and above were significantly correlated with decreases in the mycorrhizal colonisation of linseed (Seymour 2002) and wheat (Thompson 1990) in Vertosols from the Darling Downs.

Herbicides

Much of the research on herbicide effects on soil biota has been done overseas but our increase in reliance on herbicides in zero-till systems in Australia has no doubt had impacts on soil biota. Glyphosate has stimulated populations of fungi and actinomycetes with general increases in overall microbial activity even though bacterial populations were reduced (Araujo *et al.* 2003). Glyphosate and chlorosulfuron have each been associated with increased levels of Pythium root rot in barley seedlings (Blowes 1987), and take-all fungus (Mekwatanakarn and Sivasithamparam 1987). The application of chlorosulfuron increased root disease by *Rhizoctonia solani* but had no effect on take-all levels (Rovira and McDonald 1986).

Some other herbicide effects are listed below in Table 1. Generally, herbicides don't appear to cause major changes on populations of earthworms while smaller organisms can be adversely affected.

Fungicides

If a fungicide improves crop growth though eliminating foliar or soil-borne diseases, then increased organic matter in the system will generally boost microbial activity. Fungicides may however have direct effects on the non-target organisms particularly the saprobic and symbiotic soil-borne fungi. Benomyl for instance is particularly toxic to mycorrhizal fungi (Smith *et al.* 2000) which could have implications for the nutrition of the plants. Foliar-applied sprays that miss the target leaves and spray drift could also cause undesired/unintentional impacts on soil biota.

Conclusions

There is clear evidence that non-target soil biota are influenced by pesticides and fertilisers but the impacts are wide ranging – some are stimulatory, others highly inhibitory. Effects may be direct or indirect and are dependent upon several interacting factors that relate to the mode of application, the soil environment and the availability of food sources at the time of sampling. The dynamic nature of soil biology, and the effects of environment on the fate of chemical or fertiliser added and on the populations of different functional groups of biota, makes it very difficult to draw conclusions about the impacts of various inputs in our agricultural systems.

Fertilisers generally increase plant production and hence organic matter levels in soil are increased. This is generally beneficial for soil biota as food sources are increased. Some direct

toxic effects of fertilisers on microorganisms have been reported but generally effects are only in the band of application and very high levels of the chemical are required to cause damage.

Pesticides depending on their purpose may have a positive (insecticide or fungicide allow increased plant production) or negative (herbicide will decrease organic matter inputs from weeds) but may also have direct impact on non-target soil biota. In broad-acre agriculture, soil-applied pesticides (eg broad-spectrum fumigants, nematicides, fungicidal drenches) are generally uneconomic and therefore not often used. Particular pesticides may affect short-life cycle organisms such as bacteria, but recover in relatively short time frames and so the productivity of the cropping system may not have been influenced at all. However, populations of longer-lived organisms such as omnivorous nematodes and earthworms take longer to recover from damage and therefore effects are more likely to be seen in the longer term.

In a review of the impacts of pesticides on soil biota by Van Zwieten (2004), it is noted that the long-term effects or significance for future health of agroecosystems in Australia are still unknown as few studies have been conducted over several cropping seasons. Research on the implications of agricultural management on soil biology needs to continue in order to better understand the interactions between crop management and soil biology.

Table 1. Some other examples of impacts of pesticides and fertilisers reported in scientific literature.

Chemical or product	Organism/s	Effect or impact on non-target microorganisms	Reference
Pesticides			
Herbicides	Nitrifying and denitrifying bacteria	Prosulfuron inhibited N ₂ O and NO production by the bacteria	(Kinney <i>et al.</i> 2005)
	Mycorrhizal fungi	Decreased in some situations	(Dodd and Jeffries 1989)
	Protozoa	Reductions due to recommended rates of 2,4-D, simazine, diuron, monuron, cotoran. Increases in protozoa attributed to stimulation of bacteria and fungal populations as herbicide is decomposed	See (Gupta 1994)
	Earthworms	No effect in top 10cm	(Mele and Carter 1999)
	Microarthropods and microflora	Paraquat and glyphosate altered activities and reduced decomposition of crop residues.	(Hendrix and Parmelee 1985)
	Collembola and mites	Adverse effects of atrazine and simazine for up to four weeks	See (Gupta 1994)
Insecticides	Bacteria	Chlorpyrifos reduced numbers	(Pandey and Singh 2004)
	Fungi	Chlorpyrifos significantly increased numbers	(Pandey and Singh 2004)
	Protozoa	Diazinon decreased protozoan populations	(Ingham and Coleman 1984)
	Earthworms	Extremely sensitive to organophosphates and carbamates, less sensitive to organochlorines although can be affected over time due to persistent nature of these chemicals	See (Fraser 1994)
Fungicides	Nitrifying and denitrifying bacteria	Mancozeb and chlorothalonil inhibited N ₂ O and NO production	(Kinney <i>et al.</i> 2005)
	Earthworms	Copper oxychloride used in orchards is very toxic	(Lee 1985)
Fertilisers			
Phosphorus	Mycorrhizal fungi	Several reports that increasing P concentration to very high levels decreases colonisation of roots and/or spore numbers in soil	(Jensen and Jakobsen 1980) (Seymour 2002) (Hayman <i>et al.</i> 1975)
	Earthworms	Increased numbers with increasing P applications to pasture probably due to increased plant productivity	Fraser <i>et al.</i> 1993

Chemical or product	Organism/s	Effect or impact on non-target microorganisms	Reference
Nitrogen	Mycorrhizal fungi	Spore number and root colonisation decreased	(Hayman 1970)
		Effect varied with site	(Jensen and Jakobsen 1980)
	Protozoa	Significant increases, stabilisation and decreases all reported depending on situation	See (Gupta 1994)
	Actinomycetes	No effect on total counts	(Zaitlin <i>et al.</i> 2004)
	Earthworms	Increases due to long-term applications to wheat at Rothamstead (130 yrs), barley (5yrs)	See (Fraser 1994)
	Root-lesion nematodes	<i>Pratylenchus thornei</i> increased with long-term use of N fertiliser of wheat crops.	(Thompson 1992)
Lime	Mycorrhizal fungi	Little effect on colonisation; change in fungi present	(Wang <i>et al.</i> 1985)
	Earthworms	Often increases populations – response due to pH change rather than increased calcium supply	See (Fraser 1994)
Sulfur	Bacterial-feeding protozoa	30-71% decline in populations	(Gupta and Germida 1988)
	Fungal-feeding amoebae	>84% decline in fungal-feeding amoebae	(Gupta and Germida 1988)
	Fungi	reduced biomass	

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